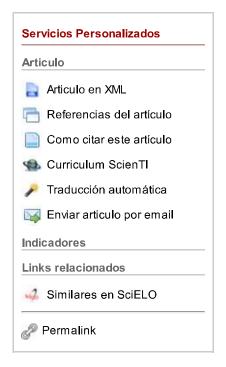




Journal of the Chilean Chemical Society versión On-line ISSN 0717-9707

J. Chil. Chem. Soc. v.51 n.2 Concepción jun. 2006

http://dx.doi.org/10.4067/S0717-97072006000200001



J. Chil. Chem. Soc., 51, No 2 (2006), pags: 837-841

ALKALOIDS AND A FLAVONOID FROM AERIAL PARTS (LEAVES AND TWIGS) OF DUGUETIA FURFURACEA - ANNONACEAE

Carlos Alexandre Carollo, Aline Regina Hellmann-Carollo, João Máximo de Siqueira* Sérgio Albuquerque

Laboratório de Farmacognosia, Departamento de Farmácia-Bioquímica, CCBS, Universidade Federal de Mato Grosso do Sul, C.P. 549, Campo Grande, MS 79070-900, Brazil

Departamento de Análises Clínicas, Toxicológicas e Bromatológicas Faculdade de Ciências Farmacêuticas de Ribeirão Preto, Universidade de São Paulo, Avenida do Café s/n, 14040-903 Ribeirão Preto, SP, Brazil

ABSTRACT

Duguetia furfuracea is a shrub of the Annonaceae family that occurs in several regions in Brazil. In the state of Mato Grosso do Sul this species often becomes an invading plant when the cerrado is turned into pastures. In the local folk medicine, its powdered seeds are mixed with water and used against

pediculosis, and the infusion of twigs and leaves is used to treat rheumatism. The present work led to the isolation and characterization from the aerial parts of the plant (leaves and twigs), of twelve isoquinoline alkaloids of aporphine oxoaporphine, bisbenzyltetrahydro-isoquinoline, benzyltetrahydro-isoquinoline and tetrahydroprotoberberine skeletons. In addition b-sytosterol and the flavonoid isorhamnethin were obtained. The activity of these different patterns of isoquinoline skeleton and the flavonoid was tested against *Trypanosoma cruzi*, and only aporphinoid skeleton with (*R*)-configuration, (-

14/2/2014 Journal of the Chilean Chemical Society - ALKALOIDS AND A FLAVONOID FROM AERIAL PARTS (LEAVES AND TWIGS) OF DUGUETIA FURFU...
)-asimilobine, was really effective against trypomastigotes (strain Y), killing approximately 72% of the parasites at a maximal concentration of 128.0 mM (IC₅₀ 57.2 mM). The trypanocidal activity of the alkaloids and the flavonoid have been investigated.

Keywords: Duguetia furfuracea, Annonaceae, alkaloid, flavonoid, Trypanosoma cruzi.

INTRODUCTION

Duguetia furfuracea is a shrub of the Annonaceae family that occurs in several regions in Brazil. In the state of Mato Grosso do Sul this species often becomes an invading plant when the cerrado is turned into pastures¹⁾. In the local folk medicine, its powdered seeds are mixed with water and used against pediculosis^{2,3)}, and the infusion of twigs and leaves is used to treat rheumatism⁴⁾. Although some chemical data are available for Brazilian native species of this genus, few biological^{5,6)} and chemical studies have been conducted with *D. furfuracea*. These include reports of the isolation, from the leaves of the plant, of flavonoid glycosides identified by HPLC-MS-ES and UV and visible spectrometric data⁷⁾ and of several other sesquiterpenoids obtained from the volatile oil⁸⁾. Antiplasmodial⁵⁾ and protozoal⁶⁾ activities have also been reported, and capsules containing "*D. furfuracea* material" indicated for renal disorders are currently being patented⁹.

The present paper describes the isolation and characterization of 12 alkaloids from aporphine, oxoaporphine, benzyltetrahydro-isoquinoline, bisbenzyltetrahydro-isoquinoline, and tetrahydroprotoberberine skeletons. In addition, b-sytosterol and the flavonoid isorhamnetin were isolated from different extracts obtained from the aerial parts (leaves and twigs) of the plant. The activity of these isoquinoline alkaloids and of the flavonoid was tested against Tripomastigote forms of Y strain protozoan parasite *Trypanosoma cruzi*.

EXPERIMENTAL

General experimental procedures

Melting points were determined on a Uniscience 498 apparatus and remain uncorrected. Optical rotations were run in chloroform on a Perkin Elmer 341 instrument. All NMR experiments were performed on a DPX-300 Brucker instrument (1 H: 300 MHz; 13 C: 75 MHz) using CDCl $_{3}$ as solvent and TMS as the internal standard. Chemical shifts are reported in d units and coupling constants (1) in Hz. 60 G and 60 GF $_{254}$ silica gel (Merck) and 70-230 mesh (Aldrich) were used for TLC and CC, respectively. TLC impregnated with 2% or 5% oxalic acid was also performed, using the usual methodology 10 . (Caution! Oxalic acid releases toxic vapors when heated.)

Plant material

The aerial parts (leaves and twigs) of *Duguetia furfuracea* (A. St.-Hil.) Benth. & Hook. f. Annonaceae, were collected on the UFMS campus in Campo Grande, MS, Brazil, and identified by Prof. R. Mello Silva. A voucher specimen (number 023) has been deposited in the CGMS Herbarium (Campo Grande, MS).

Extraction and isolation

Preparation and fractionation of the hexane extract from leaves

Powdered dried leaves of *D. furfuracea* (160 g) were submitted to exhaustive hexane extraction in a soxhlet apparatus and yielded 5.3 g of hexane extract. By submitting 4.5 g of the extract to column chromatography (150 g of silica-gel) and using hexane, methylene chloride and methanol as eluents, b-sytosterol was isolated.

Preparation of the ethanol extract from the leaves

4.8 kg of powdered dried leaves were submitted to percolation in 95% ethanol for 72 h, furnishing 0.97 kg of extract after concentration.

Partitioning of the ethanol extract obtained from the leaves

The ethanol extract (0.97 kg) was shaken for 6 h with a 3% HCl solution. The marc was discarded and the resulting acidic layer (1) was partitioned with methylene chloride, furnishing a methylene chloride layer (A) and an acidic (2) layer. Layer A was washed with water and, from the organic solvent,

and then extracted with methylene chloride, furnishing a fraction (14 g) that was labeled "DfFalk-a", which was then solubilized in chloroform and submitted to partition with 3% HCl, yielding acidic (3) and chloroform layers. The acidic (3) layer was submitted to a 10% NH₄OH/CHCl₃ procedure and furnished an alkaloid fraction labeled "DfFalk-b," which was fractionated as described below. The chloroform layer mentioned above was sequentially extracted with 3% HCl, 10% NH₄OH and CHCl₃, and the resulting chloroform layer was labeled "DfFalk-c".

Further partitioning of the "DfFalk-b" fraction

This fraction (6 g) was submitted to column chromatography with aluminum oxide and a hexane:chloroform:methanol gradient, yielding an aterospermidine-liriodenine mixture (10/11), noriscorydine (2), xylopine (3), an obovanine-anonaine mixture (4/5), ()-asimilobine (6), isochondodendrine (7), ()-discretamine (8) and (+)-reticuline (9). The aterospermidine-liriodenine fraction (15 mg) was submitted to column chromatography on silica gel (25 g) and a chloroform: methanol gradient plus 0.5% conc. HCl, allowing each alkaloid to be separated.

Further partitioning of the "DfFalk-c" fraction

This fraction (2 g) was submitted to a column chromatography on silica gel (200 g) and a chloroform: methanol gradient, yielding stricto sensu alkaloid (+)-isocorydine (1) and a complex mixture of alkaloid fractions that has been not worked yet .

Preparation and fractionation of the alkaloid extract obtained from the bark of twigs

500 g of powdered dried bark of twigs were macerated with 10% ammonium hydroxide and submitted to exhaustive extraction with methylene chloride for 15 days, yielding 10.5 g of extract. By submitting this methylene chloride extract to the usual alkaloid extraction, 1.4 q of alkaloid extract was obtained, which was then fractionated by column chromatography with aluminum oxide (250 g) and an ethyl acetate: methanol: water gradient, yielding a fraction that contained a mixture of aterospermidine, liriodenine and lanuginosine.

This fraction (52 mg) was submitted to preparative TLC on silica gel impregnated with 2% oxalic acid and eluted four times in chloroform: methanol 90:10, yielding fractions A (12 mg) and B (20 mg). Fraction A contained atherospermidin (3) mainly, whereas fraction B included two oxoaporphine alkaloids. The latter fraction was again submitted to preparative TLC impregnated with 5% oxalic acid, using methylene chloride: methanol 83:17 as eluent, and yielded fractions B' and B". In fraction B' (8 mg) it was possible to confirm the presence of liriodenine (11), predominantly; from fraction B", lanuginosine (12) was isolated.

Identification of the isolated compounds

(+)-Isocorydine (1): chloroform soluble crystals, 50 mg (0.001%) from bark alkaloid extract, optical rotation, melting point ¹⁹⁾, as well ¹H and ¹³C MNR data¹¹⁾ are in agreement with those reported in the literature.

-Norisocorydine (2): dark brown, amorphous, chloroform soluble, 10 mg (0.00021%) from bark alkaloid extract. ¹H and ¹³C MNR data are in agreement with those reported in the literature ¹²).

Xylopine (3): light brown, amorphous, chloroform soluble, 5 mg (0.0001%) from bark alkaloid extract. The 1 H NMR spectral data was comparable with those described in the literature $^{13)}$.

Obovanine-anonaine mixture (4/5): Amorphous, chloroform soluble, 30 mg (0.00063%) from bark alkaloid extract. The ¹H and ¹³C NMR spectral data from the mixture were comparable with those found in the literature for each substance separately 14,15).

(—)-Asimilobine (6): Dark brown, amorphous, chloroform soluble, 40 mg (0.00083%) from bark alkaloid extract. The ¹H and ¹³C NMR spectral data and optical rotation were comparable with those described in the literature $^{16,17)}$.

Isochondodendrine (7)¹⁸); brown, amorphous, slightly soluble in chloroform, soluble in chloroform; methanol 9:1 and chloroform: DMSO 1:1. 32 mg (0.00067%) from bark alkaloid extract. MS-ESI(+) m/z (rel. int.): 596 [M + H]⁺ (75), 299 [M/2 + H]⁺ (100). ¹H MNR (300 MHz, CHCl₃: DMSO- d_6 (1:1), δ): 2.39 Journal of the Chilean Chemical Society - ALKALOIDS AND A FLAVONOID FROM AERIAL PARTS (LEAVES AND TWIGS) OF DUGUETIA FURFU... (dd, f: 11.6; 12.4 Hz, H- α /H- α '), 2.39 (s, N-Me), 2.70-3.30 (m, H-4/H-4'/H-3/H-3'), 3.05 (br d, f: 12.8 Hz, H- α /H- α '), 3.73 (s, O-Me), 4.10 (br d, f: 10.3 Hz, H-1/H-1'), 5.63 (dd, f: 2.5; 8.5 Hz, H-11/H-11'), 6.17 (dd, f: 1.6; 8.3 Hz, H-10/H-10'), 6.43 (dd, f: 2.5; 8.2 Hz, H-13/H-13), 6.53 (s, H-5/H-5'), 6.91 (dd, f: 1.6; 8.3 Hz, H-14/H-14'), 8.08 (s, OH-7). $\frac{13}{2}$ C NMR (75 MHz, CHCl3:DMSO- $\frac{1}{2}$ 6 (1:1), $\frac{1}{2}$ 0): 23.6 (C- $\frac{1}{2}$ 7), 38.3 (C- $\frac{1}{2}$ 7), 42.0 (N-CH3), 44.2 (C- $\frac{3}{2}$ 7), 55.7 (OMe), 59.0 (C- $\frac{1}{2}$ 7), 108.4 (C- $\frac{5}{2}$ 7), 113.6 (C- $\frac{1}{2}$ 7), 117.1 (C- $\frac{1}{2}$ 7), 123.9 (C- $\frac{4}{2}$ 7), 124.7 (C- $\frac{8}{2}$ 7), 127.7 (C- $\frac{1}{2}$ 9), 128.3 (C- $\frac{1}{2}$ 7), 129.7 (C- $\frac{9}{2}$ 9), 135.6 (C- $\frac{7}{2}$ 7), 137.9 (C- $\frac{8}{2}$ 8), 147.0 (C- $\frac{6}{2}$ 6), 153.9 (C- $\frac{1}{2}$ 7).

Obtaining a diacetylated derivative of isochondodendrine (7a)

12 mg of isochondodendrine were mixed with 1 mL of acetic anhydride and 1 mL of pyridine and maintained at room temperature overnight. The usual workup afforded 10 mg of a diacetylated derivative of isochondodendrine. Dark brown, amorphous, soluble in chloroform. $^1{\rm H}$ NMR (300 MHz, CDCl3, δ): 1.57 (s, Ac-Me), 2.47 (s, N-Me), 2.57 (dd, f: 12.0 Hz, H- α /H- $^{'}$), 2.90-3.49 (m, H-4/H-4' and H-3/H-3'), 3.37 (br d, f: 12.7 Hz, H- α /H- α), 3.77 (s, O-Me), 4.42 (br d, f: 10.3 Hz, H-1/H-1'), 5.78 (dd, J: 2.5; 8.3 Hz, H-11/H-11'), 6.28 (br d, f: 7.4 Hz, H-10/H-10'), 6.56 (dd, f: 2.5; 8.5 Hz, H-13/H-13), 6.60 (s, H-5/H-5'), 7.13 (br d, f: 7.4 Hz, H-14/H-14'). $^{13}{\rm C}$ NMR (75 MHz, CDCl3, d): 19.8 (Ac-Me), 23.2 (C-4/C-4'), 40.1 (C- α /C- α '), 41.0 (N-CH3), 43.2(C-3/C-3'), 56.0 (OMe), 59.4(C-1/C1'), 108.7(C-5/C-5'), 114.0 (C-11/C-11'), 117.9 (C-13/C-13'), 121.9 (C-8a/C-8a'), 128.3 (C-10/C-10'), 128.8 (C-14/C-14'), 129.8 (C-7/C-7'), 130.0 (C-9/C-9'), 131.5 (C-4a/C-4a'), 143.6 (C-8/C-8'), 151.5(C-6/C-6'), 153.7 (C-12/C-12'), 167.2 (C=0).

(+)-Discretamine (8): colorless crystals, slightly soluble in chloroform, soluble in DMSO, ethanol, 15 mg (0.00031%) from bark alkaloid extract. Optical rotation) 19 , melting point 19,20 , and 1 H and 13 C NMR data were comparable with those described in the literature.

(+)-Reticuline (9): green, amorphous, soluble in chloroform, 96 mg (0.002%) from bark alkaloid extract. [a] $_{\rm D}^{20}$: +70(CHCl $_{\rm 3}$, c: 0.0035); literature: +90 (CHCl $_{\rm 3}$, c: 0.04) 12 . 1 H and 13 C MNR data are in agreement with those reported in the literature 21).

Aterospermidine ($\mathbf{10}$): orange crystals, slightly soluble in chloroform, soluble in chloroform-methanol mixture, 12 mg, 0.001% from bark alkaloid extract, 3 mg, 0.000063% from leaf alkaloid extract. 1 H and 13 C MNR data are in agreement with those reported in the literature 22).

Liriodenine (11): Yellow crystals, slightly soluble in chloroform, soluble in methanol; obtained from the bark alkaloid extract, 8 mg, 0.00067%, and from leaf alkaloid extract, 7 mg, 0.00015%. The spectral data were comparable with those described in the literature²³⁾.

Lanuginosine (12): Dark yellow crystals, slightly soluble in chloroform, soluble in methanol, 4 mg, 0.00033% from bark alkaloid extract. The spectral data were comparable to those described in the literature??

Isorhamnetin (**13**): Light yellow, amorphous, slightly soluble in chloroform, soluble in DMSO, 30 mg (0.00063%) from leaf ethanolic extract. 1 H MNR (300 MHz, DMSO- d_6 , δ) 24 : 3.83 (s, OMe-3'), 6.19 (br s, H-6). 6.47 (br s, H-8), 6.93 (d, f: 8.2 Hz, H-5'), 7.67 (br d, f: 8.2 Hz, H-6'), 7.74 (br s, H-2'), 9.41 (s, OH-3), 9.75 (s, OH-4'), 10.81 (s, OH-7), 12.44 (s, OH-5). 13 C MNR (75 MHz, DMSO- d_6 , d): 55.8 (O-Me-3'), 93.6 (C-8), 98.2 (C-6), 103.0 (C-10), 111.8 (C-2'), 115.6 (C-5'), 121.7 (C-6'), 122.0 (C-1'), 135.8 (C-3), 146.7 (C-2), 147.4 (C-3'), 148.8 (C-4'), 156.2 (C-9), 160.7 (C-5), 163.9 (C-7), 175.9 (C-4).

In vitro anti-Trypanosoma activity

Parasite

For the trypanocidal assay, trypomastigote forms of the Y strain of T. cruzi were cultivated in LLC-MK2 cells. The cells were cultivated in RPMI medium supplemented with 2mM of L-glutamine, 10mM of NaHCO3, 100U/mL of penicillin, 100 μ g/mL of streptomycin and 5% of inactivated, at 37°C in an environment containing 5% of CO2 and 95% of humidity. The trypomastigote forms were added into the culture at 3:1 proportion. After seven days, the supernatant was removed and centrifuged furnishing free

14/2/2014 Journal of the Chilean Chemical Society- ALKALOIDS AND A FLAVONOID FROM AERIAL PARTS (LEAVES AND TWIGS) OF DUGUETIA FURFU... trypomastigote forms of the parasite for the bioassay.

Anti-trypanosoma activity

Stock solutions were prepared by dissolving the compounds in pure dimethylsulfoxide (DMSO), to obtain a final concentration of 20 mM of each compound. Aliquots of the stock solution were added to the parasite suspension, to furnish final concentrations of 8, 32 and 128 mM, respectively.

Bioassay

In a 96 well microtitle plate about 106 forms of the parasite were added, plus the compounds to be assayed. After 24 hours of incubation, the activity was evaluated by using a colorimetric technique at 595nm by the addition of a tetrazolium salt [MTT; 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide]. Also, negative controls (solvent plus culture medium) and positive control (gentian violet at 250 µg/mL) were run in parallel. All the assays were undertaken in triplicate.

RESULTS AND DISCUSSION

In order to obtain an alkaloid extract from the ethanol extract of the leaves, acid-base extraction was performed. During this procedure, precipitation of isorhamnetin (13), a flavonoid, occurred in the supernatant (organic phase). This flavonoid might be an artifact, since it had previously been isolated⁷⁾ from hydrolyzed isorhamnetin glycosides present in the extract. This possibility, however, was discarded on the basis of evidence provided by TLC of the initial extract.

For this compound, ^1H and ^{13}C NMR spectral data were used to confirm the presence of a flavonoid skeleton and, together with 2D NMR (HMQC and HMBC) experiments, they provided support for the determination of the methoxyl group position. A singlet signal at δ 3.83 (δ_{C} 55.8) indicated the presence of a methoxyl group, corresponding to a group linked to C-4' in ring B.

Spectral methods in combination with optical activity data provide an useful tool to help elucidate aporphine sensu stricto alkaloid skeletons $^{12,13)}$. Six already known aporphine sensu stricto alkaloids obtained from the bark of twigs namely, (+)-isocorydine (1), norisocorydine (2), xylopine (3), obovanine

(4), anonaine (5), and ()-asimilobine (6) were thus characterized by comparing their 1 H and 13 C spectral data with those available in the literature 18).

Isochondodendrine (**7**), a dimeric aporphinoid alkaloid with a bisbenzyltetrahydro-isoquinoline skeleton, was also isolated. MS-ESI(+) revealed a pseudomolecular ion at m/z 596 ([M + H]⁺) and a fragment ion at m/z 299 ([M/2 + H]⁺). The ¹H and ¹³C NMR spectral data for **7**, as well as those of its diacetylated derivative, were in accordance with those described in the literature¹³.

Compound ${\bf 8}$ was isolated as colorless crystals (acetone) and showed positive reaction to Dragendorff's reagent. After uni- and bidimensional analysis of the NMR spectral data and a negative specific rotation value it was possible to confirm ${\bf 8}$ as ()-discretamine, a tetrahydro-protoberberinic alkaloid. By comparison of its $^1{\rm H}$ and $^{13}{\rm C}$ NMR spectral data with those described in the literature, some assignments of carbon resonance were corrected.

Compound $\mathbf{9}$, on the basis of its specific rotation and spectral (1 H and 13 C NMR) data, which were in accordance with those available in the literature, was identified as being (+)-reticuline, a benzyltetrahydroisoquinolinic alkaloid.

(1) $R_1 = R_2 = R_5 = OCH_3$, $R_3 = R_4 = H$, $R_6 = OH$, $R_7 = CH_3$, and $6a \alpha$;

(2) $R_1 = R_2 = R_5 = OCH_3$, $R_3 = R_4 = H$, $R_6 = OH$, $R_7 = H$, and 6a

(3) $R_1 + R_2 = OCH_2O$, $R_4 = OCH_3$, $R_3 = R_5 = R_6 = R_7 = H$, and 6a α :

(4) $R_1 + R_2 = OCH_2O$, $R_6 = OH$, $R_3 = R_4 = R_5 = R_7 = H$, and $6a \alpha$;

(5) $R_1 + R_2 = OCH_2O$, $R_3 = R_4 = R_5 = R_6 = R_7 = H$, and 6a α ;

(6) $R_1 = OCH_3$, $R_2 = OH$, $R_3 = R_4 = R_5 = R_6 = R_7 = H$, and $6a \beta$.

Anti-trypanosoma activity

Alkaloidal and non-alkaloidal extracts and isolated compounds from *Duguetia* genus, as well as another species of Annonaceae family were previously tested for *in vitro* antiplasmodial and antiprotozoal

activities. In general, the alkaloid extracts show more activity than others⁶⁾ and those activity has been related with the presence of isoquinoline alkaloid-type. While the antiplasmodial activity of some alkaloids isolated from D. vallicola belong to oxoaporphinoids, (-)-7-hydroxy-aporphinoids and aza-antraquinone (cleistopholine), only the isoquinoline alkaloid, (-)-oliveroline was really effective, followed by cleistopholine²⁵. In the present work, as reported on Table 1, the lysis percentages of T. cruzi trypomastigote forms, represents the trypanocidal activity of the compounds isolated $\mathbf{1}$, $\mathbf{6}$, $\mathbf{7}$, $\mathbf{8}$, $\mathbf{9}$, $\mathbf{10}/\mathbf{11}$ mixture and $\mathbf{13}$. Only aporphinoid skeleton with (R)-configuration, (-)-asimilobine, ($\mathbf{6}$) was also effective, killing approximately 72% of the parasites at maximal concetration of 128.0 mM (IC₅₀ 57.2 mM). The present result could, at least, provide supports for the structure activity relationship between this kind of aporphinoid alkaloids and the activity in subject.

Table 1. Percentual of lysis activity against T. cruzy trypomastiqote form (strain Y)

Compound -	Concentration in μM and percentual of parasitary lise (± SD)			IC ₅₀ (μΜ)
	8.0	32.0	128.0	
1	4.9 ± 0	16.1 ± 0	40.6 ± 4.9	193.5
6	0.7 ± 0.3	35.7 ± 3.9	72.0 ± 7.9	57.2
7	12.6 ± 4.9	19.6 ± 4.9	35.7 ± 1.9	418.4
9	7.0 ± 3.0	9.1 ± 5.9	34.3 ± 5.9	252.8
8	19.6 ± 3.0	24.5 ± 4.0	35.7 ± 3.9	945.2
0/11 mixture	2.1 ± 0	18.9 ± 3.9	19.6 ± 0	-
13	4.2 ± 1.9	4.9 ± 0	12.6 ± 3.0	4.635,0

Positive control gentian Violet to 250 μ g/mL (IC₅₀ = 31 μ M); Negative control Mice infected blood + DMSO;

ACKNOWLEDGEMENTS

The authors are grateful to FUNDECT-MS, Brazil, for the financial support and to PIBIC-CPq-PROPP-UFMS and FUNDECT-CNPq for the fellowship granted. Thanks are also given to FCFRP-USP-SP for making their facilities available for additional analyses.

REFERENCES

- 1. H. Lorenzi, *Plantas daninhas do Brasil*, 3^a ed., Ed. Plantarum, Nova Odessa, SP. 2000. [Links]
- 2. M. P. Correa, *Dicionário de Plantas Úteis do Brasil e das exóticas cultivadas*, 6 vol., II ed., Imprensa http://www.scielo.cl/scielo.php?script=sci_arttext&pid=S0717-97072006000200001&lang=pt

Nacional, Rio de Janeiro, 1978. [Links]

- 3. I. Silberbauer-Gottsberger, Oreades 14/15, 15 (1981/82). [Links]
- 4. V. E. G. Rodrigues, D. A. Carvalho, Ciên. agrotec. 25, 102 (2001). [Links]
- 5. D. C. H. Fischer, N. C. A. Gualda, D. Bachiega, C. S. Carvalho, F. N. Lupo, S. V. Bonotto, M. O., Alves, A. Yogi, S. M. Santi, P.E. Ávila, K. Kirchgatter, P. R. H. Moreno, Acta Tropica 92, 261 (2004).

 [Links]
- 6. A. G. Tempone, S. E. Treiger Borborema, H. F. de Andrade Jr., N. C. de Amorim Gualda, A. Yogi, C. Salerno Carvalho, D. Bachiega, F. N. Lupo, S. V. Bonotto, D. C. H. Fischer, Phytomedicine 12, 382 (2005). [Links]
- 7. D. Y. A. Santos, M. L. F. Salatino, Phytochemistry 55, 567 (2000). [Links]
- 8. C. A. Carollo, A. R. Hellmann, J. M. de Siqueira, Bioch. Syst. Ecol. 33, 647 (2005). [Links]
- 9. L. Da Silva Coelho, Dervewent Innonvations Index Br20022030- A, (2003). [Links]
- 10. D. B da Silva, H. S. Miglio, C. A. Carollo, J. R. Fabri, J. M. de Siqueira, 28a Reunião Anual da Sociedade Brasileira de Química, Poços de Caldas, MG, Brazil, 2005, PN 258, available in www.sbq.org.br. [Links]
- 11. M-H.Yang, A. V. Patel, G. Blunden, C. H. Tumer, M. J. O'Neill, J. A. Lewis, Phytochemistry 33, 943 (1993). [Links]
- 12. A. Cavé, M. Leboeuf, P. G. Watermann, *The Aporphinoid alkaloids of the Annonaceae In Chemical and Biological perspectives, Ed. S. Willian Pelleiter, v. 5, New York, 1987.* [Links]
- 13. H. Guinaudeau, M. Leboeuf, A. Cave, J. Nat. Prod. 46, 1761 (1983). [Links]
- 14. H. Guinaudeau, M. Leboeuf, A. Cave, J. Nat. Prod. 51, 1033 (1994). [Links]
- 15. V. R. Navarro, I. M. F. Sette, D. V. L. da Cunha, M. S. Silva, J. M. Barbosa-Filho, J. G. S., Rev. Bras. Pl. Med. 3, 23 (2001). [<u>Links</u>]
- 16. T. J. Hsieh, F-R. Chang, Y-C. Wu, C-P. Cho, J. Chin. Chem. Soc. 46, 607 (1999). [<u>Links</u>]
- 17. H. Guinaudeau, M. Leboeuf, A. Cave, J. Nat. Prod. 42, 325 (1979). [<u>Links</u>]
- 18. A. J. Marsaioli, E. A. Rúveda, F. A. M. Reis, Phytochemistry 17, 1655 (1978). [Links]
- 19. S-T. Lu, Ian-Lih; T. S-P. Leou, Phytochemistry 24, 615 (1989). [Links]
- 20. J. T. Blanchfield, D. P. A. Sands, C. H. L. Kennard, K. A. Byriel, W. Kitching, Phytochemistry 63, 711 (2003). [Links]
- 21. W. S. Garcez, Tese de Doutorado, Universidade de São Paulo, São Paulo, 1991. [Links]
- 22. I. Castro, J. R. Villegas, B. Soeder, O. Castro, Fitoterapia 57, 181 (1996). [Links]
- 23. M. Nieto, A. Cavé, M. Leboeuf, J. Nat. Prod. 39, 350 (1976). [Links]
- 24. P. K. Agrawal, R. S. Thakur, M. C. Bansal, M. C. *In P. K. Agrawal Carbon-13 NMR of Flavonoids studies in organic chemistry*, v. 39, Ed. Elsevier, New York (1989). [Links]
- 25. E. Pérez, J. Sáez, S. Blair, X. Franck, B. Figadère, Let. Org. Chem. 1, 102 (2004). [<u>Links</u>]

e-mail: <u>imaximo@nin.ufms.br</u>

P.O. Box 2613, Concepción, Chile Phone 41-2227815, Fax 41-2235819

e-Mail

schqjournal@entelchile.net